



BIOPLAST

**Laboratory of BIOlogy
and regenerative medicine-PLASTic surgery**

Direttrice Prof.ssa Adriana Cordova
Responsabile Scientifico Prof. Francesco Moschella

**ATTIVITÀ SCIENTIFICA
2018-2022**

Indice

Il laboratorio	_____	01
Attività di formazione	_____	03
Pubblicazioni	_____	06
Relazioni a convegni	_____	19
Collaborazioni	_____	23
Research team	_____	24

IL LABORATORIO

PRESENTAZIONE



La chirurgia rigenerativa rappresenta in un certo senso la chirurgia del futuro, così come preconizzato a suo tempo da Joseph Murray, chirurgo plastico e premio Nobel per la Medicina nel 1990, autore del primo trapianto di rene al mondo nel 1954. Murray l'aveva chiamata "Chirurgia induttiva", partendo dal principio di potere creare organi ex novo da utilizzare in chirurgia dei trapianti. Egli in quegli anni aveva avuto una 'visione' ed il suo intuito si è via via materializzato quando nel 1981 alcuni ricercatori riuscirono per la prima volta a coltivare cellule staminali da embrioni di topo e, successivamente, nel 1998 Thomson ha creato la prima linea al mondo di cellule staminali embrionali umane. Negli anni successivi, agli inizi del terzo millennio, si è scoperto che facendo crescere le staminali embrionali (in vitro) in un ambiente tridimensionale, queste si possono organizzare in tessuti complessi detti 'Organoidi'.

Per motivi etici la ricerca sulle Staminali Embrionali in tutto il mondo non è stata incentivata, diversamente dalle ricerche sulle staminali adulte multipotenti estraibili da vari tessuti.

E' stata determinante, specialmente per la chirurgia plastica, la scoperta che il tessuto adiposo è un vero e proprio organo dinamico coinvolto in un'ampia gamma di processi biologici e che rappresenta la fonte principale di cellule staminali mesenchimali per uso clinico e sperimentale perchè si può prelevare in grandi quantità senza lasciare esiti. Il settore della medicina e chirurgia rigenerativa, in rapida espansione,

si occupa dello sviluppo di terapie innovative che hanno la finalità di ricostruire tessuti e organi danneggiati, facendo ricorso ad una popolazione di cellule staminali adulte già presenti negli stessi tessuti e in grado di autorinnovarsi. Lo sviluppo della tecnologia ha consentito il loro impiego come terapia cellulare (es.: sono state utilizzate per la riparazione della cornea ed il recupero della funzione visiva) o come terapia genica per la riparazione di difetti genetici (es.: alcune forme di epidermolisi bollosa).

La possibilità di utilizzare bio-stampanti 3D con l'ausilio di vari tipi di bio-inchiostri (idrogel) ha aperto ai ricercatori un mondo nuovo e affascinante, impensabile fino a pochi anni addietro. Infatti, facendo ricorso alle tecnologie di bio-printing in 3D, ricercatori di tutto il mondo si stanno cimentando nella stampa di organi, pelle, cartilagine e ossa. Si tratta della creazione di strutture tri-dimensionali che allo stato attuale non hanno un immediato impiego clinico per problemi inerenti la loro vascolarizzazione e quindi la loro sopravvivenza.

Quest'ultimo aspetto (costrutti tri-dimensionali) rappresenta il settore di ricerca che stiamo portando avanti nel laboratorio BIOPLAST del Policlinico Universitario, con particolare interesse per il bio-printing di tessuto osseo e cartilagineo, portando avanti un settore affascinante e anche in un certo senso rivoluzionario in campo biomedico.

Prof. Francesco Moschella

MISSIONI

La missione principale del Laboratorio Bioplast è la ricerca scientifica.

Molteplici linee di ricerca di base e precliniche sono sviluppate in laboratorio, grazie a finanziamenti di ateneo e su fondi ministeriali.

Le principali attività scientifiche oggetto dei nostri studi vertono su due importanti ed attuali filoni di ricerca:

- 1) Medicina Rigenerativa**
- 2) Oncologia**

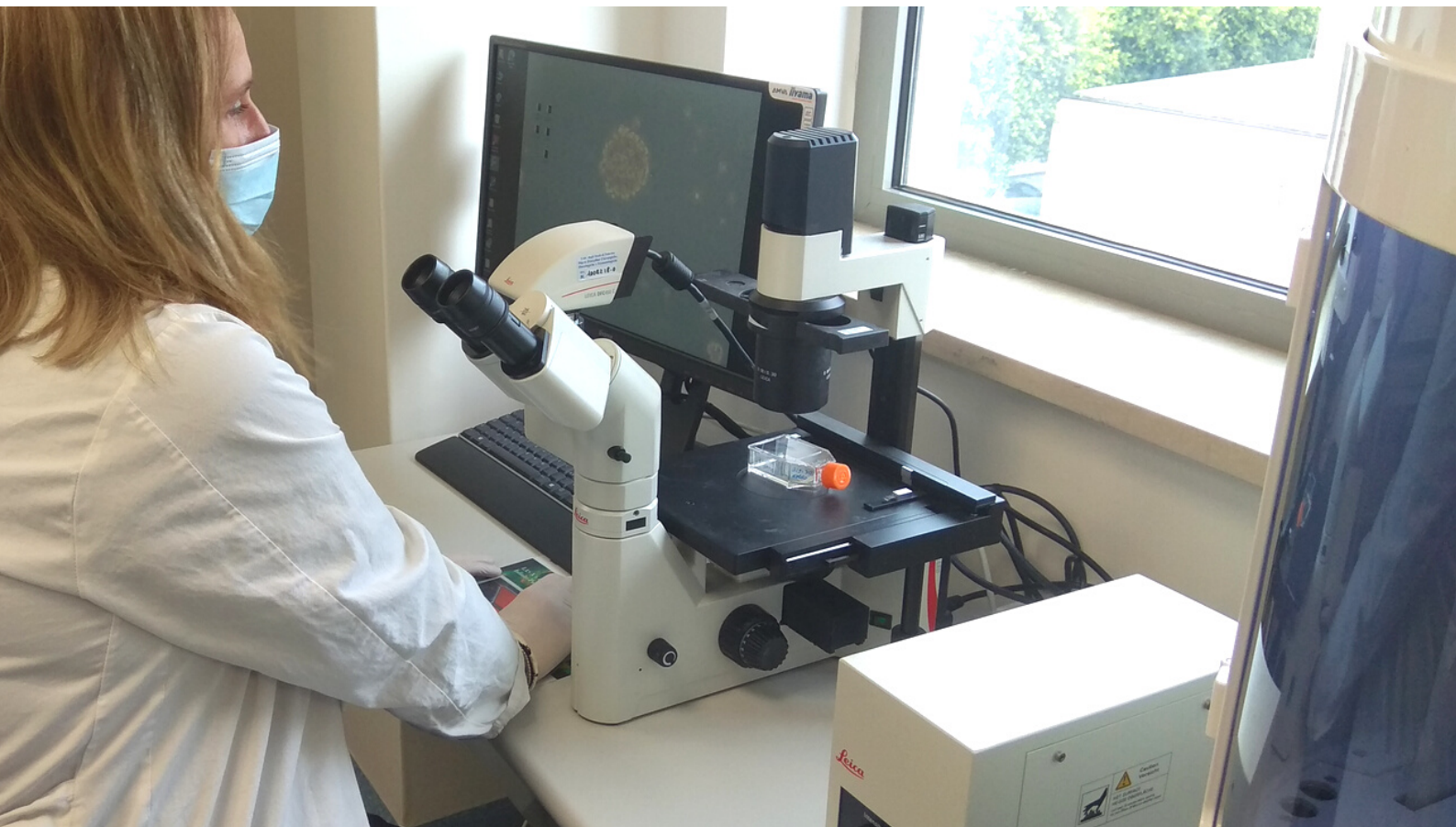
Il laboratorio Bioplast è dotato di attrezzature all'avanguardia necessarie per effettuare analisi complete a 360°.

Principali attività scientifiche:

- Studio di una nuova popolazione di **sferoidi di cellule staminali adipose** (SASCs), caratterizzazione molecolare: analisi genomica e proteomica.
- Studio di una nuova popolazione di **sferoidi di cellule staminali mesenchimali da cordone ombelicale**, caratterizzazione molecolare: analisi genomica e proteomica.
- Studio del **ruolo immunomodulatorio** di sferoidi di cellule staminali adipose nei trapianti compositi vascolarizzati: induzione della tolleranza immunologica e monitoraggio in vivo.
- 3D-printing** di nuovi bioinks biocompatibili per la ricostruzione di lesioni ossee e cartilaginee.
- Analisi e confronto tra le diverse **sedì anatomiche di prelievo** e tecniche di estrazione di cellule staminali da tessuto adiposo
- Caratterizzazione del **secretoma** prodotto dagli sferoidi di cellule staminali adipose (SASCs).
- Ricerca di **nuovi immunocheckpoints** tumorali.



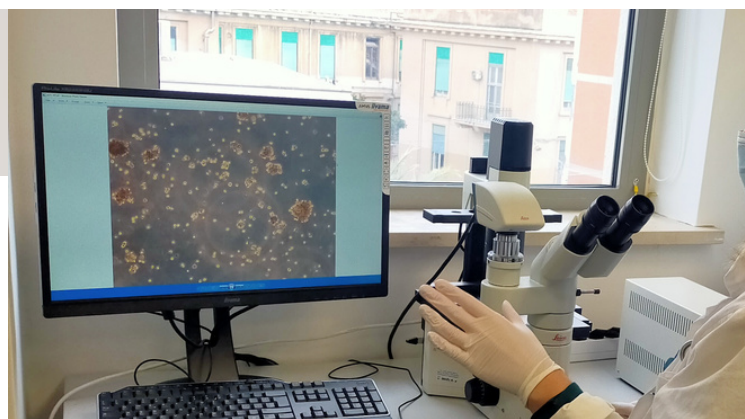
MISSIONI



Un'altra missione fondamentale del Laboratorio Bioplast è quella di fornire un adeguato **supporto alla didattica di formazione multidisciplinare** ad ampio spettro: dai corsi di laurea triennali fino a quelle specialistiche di Scienze Biologiche, Biotecnologie Mediche, Ingegneria Biomedica e Medicina e Chirurgia, alla formazione post-laurea nei corsi di dottorato e di scuola specializzazione in Chirurgia Plastica e Ricostruttiva.

Non per ultimo, il laboratorio ha in attivo numerose **collaborazioni multidisciplinari** ed è sempre disponibile ad instaurarne di nuove al fine di promuovere e implementare nuove idee di ricerca scientifica.

ATTIVITÀ DI FORMAZIONE MULTIDISCIPLINARE



Corso di Laurea Triennale in Scienze Biologiche, Dipartimento di Scienze e Tecnologie biologiche, chimiche e farmaceutiche, Università degli Studi di Palermo:

- 2022. Silvia Fontana, Tirocinio curriculare 300 ore.
Titolo della tesi: Analisi dei fattori secreti dagli sferoidi di cellule staminali adipose.
- 2022. Sofia Bulgarello, Tirocinio curriculare 300 ore.
Titolo della tesi: In corso

Corso di Laurea magistrale in Ingegneria Biomedica, Dipartimento di Ingegneria, Università degli Studi di Palermo:

- 2022. Martina Calandra, Tirocinio curriculare 150 ore
- 2022. Vincenzo Gervasi, Tirocinio curriculare 150 ore
- 2022. Maria Sofia Ferraro, Tirocinio curriculare 150 ore

Corso di Laurea magistrale II livello in Biotecnologie e Medicina Molecolare, Scuola di Medicina e Chirurgia, Dipartimento di Biomedicina, Neuroscienze e Diagnostica avanzata, Università degli Studi di Palermo:

- 2017. Federica Grisafi, Titolo della tesi: Analisi dei profili di espressione di microRNA in Sferoidi di Cellule Staminali Adipose (S-ASCs) e il loro ruolo sul mantenimento della staminalità e sul differenziamento mesenchimale.
- 2017. Riccardo Gattuccio, Titolo della tesi: Analisi molecolare di una nuova popolazione di sferoidi di cellule staminali derivate da tessuto adiposo (S-ASCs) in condizioni di "in vitro long-term culture".
- 2020. Marco Trapani, Titolo della tesi: Nuova strategia di rigenerazione tissutale mediata da sferoidi di cellule staminali adipose.
- 2021. Salvatore La Rosa, Titolo della tesi: Studio del potenziale differenziativo degli sferoidi di cellule staminali adipose.
- In corso. Emanuela Sciortino, Titolo della tesi: "Bioprinting di nuovi bioinks per la rigenerazione tissutale mediata da sferoidi di cellule staminali adipose"

Corso di Laurea Magistrale in Medicina e Chirurgia, Scuola di Medicina e Chirurgia, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Sezione di Chirurgia Plastica e Ricostruttiva, Università degli Studi di Palermo:

- 2018. Nicole Finocchiaro, Titolo della tesi: Cellule staminali da tessuto adiposo: sedi di prelievo e tecniche di manipolazione.
- 2020. Giorgio Pirrotta, Titolo della tesi: Nuovi target molecolari nell'immunoterapia del melanoma.

Scuola di specializzazione in Chirurgia Plastica, Ricostruttiva ed estetica, Scuola di Medicina e Chirurgia, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Sezione di Chirurgia Plastica e Ricostruttiva, Università degli Studi di Palermo:

- 2018. Luigi Montesano, Titolo della tesi: Studio della capacità rigenerative degli Sferoidi di Cellule Staminali derivate da tessuto adiposo (SASCs) su un difetto della calvaria in un modello animale.
- 2022. Daniele Matta, Titolo della tesi: Evaluation of the regenerative capacity of PRP-p in palate fistulas: efficacy, safety and comparison of the biochemical composition.

Dottorato di ricerca in Oncologia e Chirurgia Sperimentali, Dipartimento di Discipline Chirurgiche Oncologiche e Stomatologiche, Università degli Studi di Palermo:

- Ciclo XXXII (2016-2019). Marco Pappalardo, Titolo della tesi: Analysis of the Immunomodulatory Properties of Spheroids from Adipose-derived Stem Cells.
- Ciclo XXXIII (2017-2020). Federica Grisafi, Titolo della tesi: 3D in Suspension versus 2D in Adhesion: molecular profiles in stemness and mesenchymal differentiation of Spheroids from Adipose-derived Stem Cells.
- Ciclo XXXIV (2018-2021). Luigi Montesano, Titolo della tesi: Analysis of infiltrating gamma delta and T-reg cells in cutaneous melanoma: a hypothetic crosstalk for new immunotherapy.
- Ciclo XXXVI (2021-2024). Valentina Urrata, Titolo della tesi: Caratterizzazione del secretoma degli sferoidi di cellule staminali derivate da tessuto adiposo (S-ASCs) e loro applicazione per la rigenerazione tissutale. In corso

PUBBLICAZIONI 2018-2022

IMPACT FACTOR TOTALE: **64.345**

1

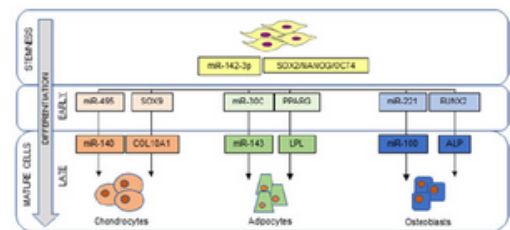
2018

Spheroids from adipose-derived stem cells exhibit an miRNA profile of highly undifferentiated cells.

Anna Barbara Di Stefano,
Federica Grisafi, Marta
Castiglia, Alessandro Perez,
Luigi Montesano, Alessandro
Gulino, Francesca Toia, Daniele
Fanale, Antonio Russo,
Francesco Moschella, Angelo A.
Leto Barone, Adriana Cordova.

J Cell Physiol. 2018
Nov;233(11):8778-8789.
doi: 10.1002/jcp.26785. Epub
2018 May 24. PMID: 29797571

I.F.: 4.522 Q1



Received: 30 November 2017 | Accepted: 30 April 2018
DOI: 10.1002/jcp.26785

ORIGINAL RESEARCH ARTICLE

WILEY Cellular Physiology

Spheroids from adipose-derived stem cells exhibit an miRNA profile of highly undifferentiated cells

A. Barbara Di Stefano PhD^{1,2} | Federica Grisafi MSc^{1,2} | Marta Castiglia PhD² |
Alessandro Perez PhD² | Luigi Montesano MD¹ | Alessandro Gulino PhD³ |
Francesca Toia MD¹ | Daniele Fanale PhD² | Antonio Russo MD² |
Francesco Moschella MD¹ | Angelo A. Leto Barone MD⁴ | Adriana Cordova MD¹

¹Department of Surgical, Oncological and Oral Sciences, Section of Plastic and Reconstructive Surgery, University of Palermo, Palermo, Italy

²Department of Surgical, Oncological and Oral Sciences, Section of Medical Oncology, University of Palermo, Palermo, Italy

³Department of Health Science, Human Pathology Section, Tumor Immunology Unit, University of Palermo, Palermo, Italy

⁴Department of Plastic and Reconstructive Surgery, Johns Hopkins School of Medicine, Baltimore, MD

Correspondence
Antonio Russo, MD, Department of Surgical, Oncological and Oral Sciences, Section of Medical Oncology, University of Palermo, via del Vespro 129, 90127 Palermo, Italy
Email: antonio.russo@unipa.it

Funding information
Italian Ministry of Health, Grant/Award Number: GR-2010-2321017

Two-dimensional (2D) cell cultures have been extensively used to investigate stem cell biology, but new insights show that the 2D model may not properly represent the potential of the tissue of origin. Conversely, three-dimensional cultures exhibit protein expression patterns and intercellular junctions that are more representative of their in vivo condition. Multiclonal cells that grow in suspension are defined as "spheroids," and we have previously demonstrated that spheroids from adipose-derived stem cells (S-ASCs) displayed enhanced regenerative capability. With the current study, we further characterized S-ASCs to further understand the molecular mechanisms underlying their stemness properties. Recent studies have shown that microRNAs (miRNAs) are involved in many cellular mechanisms, including stemness maintenance and proliferation, and adipose stem cell differentiation. Most studies have been conducted to identify a specific miRNA profile on adherent adipose stem cells, although little is still known about S-ASCs. In this study, we investigate for the first time the miRNA expression pattern in S-ASCs compared to that of ASCs, demonstrating that cell lines cultured in suspension show a typical miRNA expression profile that is closer to the one reported in induced pluripotent stem cells. Moreover, we have analyzed miRNAs that are specifically involved in two distinct moments of each differentiation, namely early and late stages of osteogenic, adipogenic, and chondrogenic lineages during long-term in vitro culture. The data reported in the current study suggest that S-ASCs have superior stemness features than the ASCs and they represent the true upstream stem cell fraction present in adipose tissue, relegating their adherent counterparts.

KEYWORDS

adipose stem cells, long-term culture, mesenchymal differentiation, miR-142-3p, miRNAs

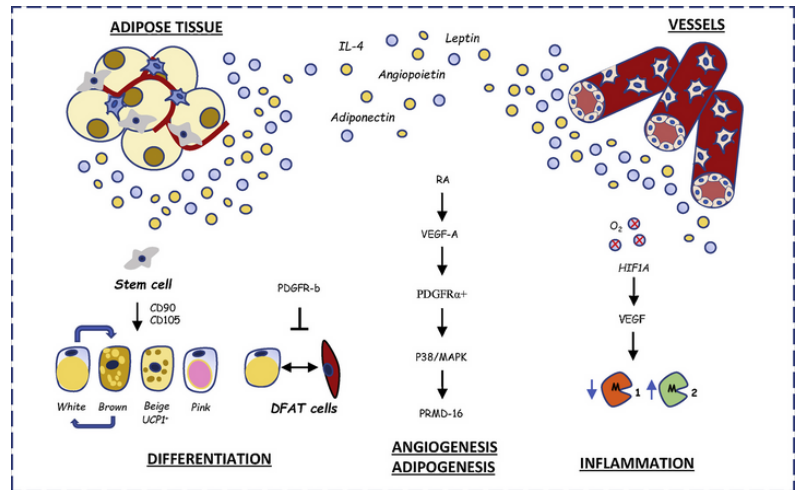
2018

Adipose tissue, angiogenesis and angio-MIR under physiological and pathological conditions.

Di Stefano AB, Massihnia D, Grisafi F, Castiglia M, Toia F, Montesano L, Russo A, Moschella F, Cordova A.
Eur

J Cell Biol. 2019 Jun;98(2-4):53-64.
doi: 10.1016/j.ejcb.2018.11.005. Epub
2018 Nov 30. PMID: 30527802

I.F.: 3.024 Q2



European Journal of Cell Biology 98 (2019) 53–64



Contents lists available at ScienceDirect

European Journal of Cell Biology

journal homepage: www.elsevier.com/locate/ejcb



Review

Adipose tissue, angiogenesis and angio-MIR under physiological and pathological conditions

Anna Barbara Di Stefano^{a,b,*}, Daniela Massihnia^{b,1}, Federica Grisafi^{a,b}, Marta Castiglia^b,
Francesca Toia^a, Luigi Montesano^a, Antonio Russo^b, Francesco Moschella^a, Adriana Cordova^a

^a Department of Surgical, Oncological and Oral Sciences, Section of Plastic and Reconstructive Surgery, University of Palermo, 90127 Palermo, Italy

^b Department of Surgical, Oncological and Oral Sciences, Section of Medical Oncology, University of Palermo, 90127 Palermo, Italy

ARTICLE INFO

Keywords:
Adipose stem cells
Endothelial cells
Angiogenesis
miRNAs

ABSTRACT

Angiogenesis is a crucial process for the maintenance of normal tissue physiology and it is involved in tissue remodeling and regeneration. This process is essential for adipose tissue maintenance. The adipose tissue is composed by different cell types including stromal vascular cells as well as adipose stem cells (ASCs). In particular, ASCs are multipotent somatic stem cells that are able to differentiate and secrete several growth factors; they are recently emerging as a new cell reservoir for novel therapies and strategies in many diseases. Several studies suggest that ASCs have peculiar properties and participate in different disease-related processes such as angiogenesis. Furthermore, pathological expansion of adipose tissue brings to hypoxia, a major condition of unhealthy angiogenesis.

Recent evidences have shown that microRNAs (miRNAs) play a crucial role also on ASCs as they take part in stemness maintenance, proliferation, and differentiation. It has been suggested that some miRNAs (MIR126, MIR31, MIR221, MIR222, MIR17-92 cluster, MIR30, MIR100 and MIR486) are directly involved in the angiogenic process by controlling multiple genes involved in this pathway. With the present review, we aim at providing an updated summary of the importance of adipose tissue under physiological and pathological conditions and of its relationship with neovascularization process. In particular, we report an overview of the most important miRNAs involved in angiogenesis focusing on ASCs. Hopefully the data presented will bring benefit in developing new therapeutic strategies.

2019

Immunomodulation in Vascularized Composite Allotransplantation: what role for Adipose-derived stem cells?

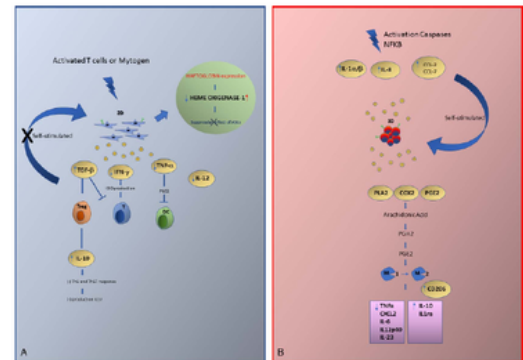
Marco Pappalardo, Luigi Montesano,
Francesca Toia, Antonio Russo,
Sara Di Lorenzo, Francesco Dieli,
Francesco Moschella, Angelo A. Leto
Barone, Serena Meraviglia,
Anna Barbara Di Stefano.

Ann Plast Surg. 2019 Feb;82(2):245-251.

doi:

10.1097/SAP.0000000000001763.PMID:

30628936



REVIEW PAPER

I.F.: 1.448 Q2

Immunomodulation in Vascularized Composite Allotransplantation What Is the Role for Adipose-Derived Stem Cells?

Marco Pappalardo, MD,* Luigi Montesano, MD,* Francesca Toia, MD, PhD,* Antonio Russo, MD, PhD,†
Sara Di Lorenzo, MD, PhD,* Francesco Dieli, MD, PhD,‡§ Francesco Moschella, MD,*
Angelo A. Leto Barone, MD,*|| Serena Meraviglia, MD,‡§ and Anna Barbara Di Stefano, PhD*†

Abstract: Hand and face transplants are becoming increasingly common, recording progressively more penis, uterus, abdominal wall, and allotransplantation cases reported worldwide. Despite current protocols allow long-term survival of the allografts, the ultimate goal of donor-specific tolerance has not been achieved yet. In fact, the harmful adverse effects related to the lifelong administration of immunosuppressive agents are the main drawbacks for vascularized composite allotransplantation. Research is very active in investigating alternative methods to induce greater tolerance while minimizing toxicity. Adipose-derived stem cells (ASCs) represent promising cell therapies for immunomodulation in pre-clinical and clinical settings. Their clinical appeal is due to their easy harvest in large quantities through a minimally invasive and well-accepted approach; they may well promote donor-specific tolerance and potentially reduce immunosuppression. Several experimental studies exist, but lacking review articles reporting current evidence. This work proposes a literature review on the immunomodulatory role of ASCs in vascularized composite allotransplantation. In vitro and in vivo evidence will be summarized. The role that cell passaging and upstream progenitors—the so-called spheroid ASCs—may play in modulating the immune response will also be discussed. Finally, this article will summarize current knowledge on biotransformation, migration, and homing of injected stem cells. This review may well provide useful information for preclinical and clinical studies, aiming at a breakthrough for donor-specific tolerance.

Key Words: adipose-derived stem cells, cell-based therapies, face transplantation, hand transplantation, vascularized composite allotransplantation

(Ann Plast Surg 2019;82: 245–251)

Since their first description, more than 100 human hand and 37 face vascularized composite allotransplants (VCAs) have been reported in the past 18 years,^{1–4} proving not only their technical feasibility, but also the superior outcomes compared with conventional reconstruction in selected patients. Nonetheless, VCA is not routinely performed because of the condition of lifelong immunosuppression and toxicity profiles.⁵ The major complications resulting from

immunosuppressive therapies include increased propensity to develop infections, organ toxicity (ie, nephrotoxicity), and malignancy.⁶

Furthermore, unlike solid organ transplantation, VCAs are life enhancing rather than lifesaving, leading to many ethical and medical issues involving individual surgeons as well as professional societies.⁷

The use of novel cell-based therapies that combine the benefits of immunomodulation with neuroregeneration proves to have a great potential in improving functional outcomes and the quality of life in VCA patients.⁸

It is known that mesenchymal stem cells (MSCs) are “immune privileged” because of their low expression of human leukocyte antigen and costimulatory molecules. Furthermore, they exert potent immunosuppressive actions.⁹ These desirable properties make MSCs a new therapeutic option; MSCs have emerged as a promising cell-based therapy for immunomodulation and are currently being examined in pre-clinical and clinical settings as therapeutic solutions for autoimmune disorders or transplant rejection.^{10–12}

To date, bone marrow-derived MSCs (BM-MSCs) are considered the standard cell type used in the fields of stem cell biology and clinical application. However, adipose-derived stem cells (ASCs), abundant in the human body, could be harvested with minimal invasiveness and are becoming increasingly widespread in treating various diseases.¹³ Many studies have aimed to investigate the phenotypic and intrinsic biological differences between traditional adherent and nonadherent ASCs.^{14,15} The recent isolation of a nonadherent ASC population paved the way to the possibility that this niche could include cells at a low advanced stage of differentiation and therefore more upstream in the stem cell hierarchy. Despite scientists have begun using spheroids from ASCs (S-ASCs) in a number of laboratories worldwide, their identity as a separate stem cell niche with different phenotype, biological behavior, and differentiation ability has never been fully defined.

The following summarizes current literature available on in vitro and in vivo studies concerning the immunomodulatory effects of ASCs and discusses the potential role that the 3-dimensional (3D) niche and upstream progenitors have in modulating the immune system. Finally, we summarize current knowledge available on biotransformation, migration, and homing of injected stem cells.

Immunomodulatory Role of ASCs

The so-called ASCs are a variety of MSCs extracted from adipose tissue and also defined as fibroblast-like, adherent, and multipotent cells. Indeed, ASCs are able to differentiate along multiple lineages including adipocytes, osteoblasts, and chondrocytes, as well as other cell types including hepatocytes, myocytes, and neuronal-like cells.^{16–17} The successful isolation of multipotent ASCs achieved in the past decade and their ability to preferentially home to damaged tissue make ASCs a promising tool in regenerative medicine.^{17,18} Indeed, ASCs can be engineered and used as biological vehicles for vector-based genes to convey therapeutic molecules in vivo.^{20,21} Furthermore,

Received February 15, 2018, and accepted for publication, after revision October 17, 2018.
From the *Plastic and Reconstructive Surgery and †Medical Oncology, Department of Surgical, Oncological and Oral Sciences, Central Laboratory of Advanced Diagnosis and Biomedical Research (CLADBIOR); and ‡Department of Biopathology and Biomedical Methodologies, University of Palermo, Palermo, Italy; and §Plastic and Reconstructive Surgery, Johns Hopkins Medicine, Department of Plastic and Reconstructive Surgery, Baltimore, MD.
Author Contributions: Special topic conceived by M.P., F.T., S.M., and A.B.D.S. Manuscript drafted by M.P., A.A.L.B., L.M., S.D.L., F.T., S.M., and A.B.D.S. Manuscript revision carried out by A.R., F.D., F.M., and A.A.L.B.
Conflicts of interest and sources of funding: none declared.
Reprints: Francesca Toia, MD, PhD, Plastic and Reconstructive Surgery, Department of Surgical, Oncological and Oral Sciences, University of Palermo, Via del Vespro, 129, 90127, Palermo, Italy. E-mail: francesca.toia@gmail.com.
Copyright © 2019 Wolters Kluwer Health, Inc. All rights reserved.
ISSN: 0148-7043/19/020245-07
DOI: 10.1097/SAP.0000000000001763

Annals of Plastic Surgery • Volume 82, Number 2, February 2019

www.annalsplasticsurgery.com | 245

2019

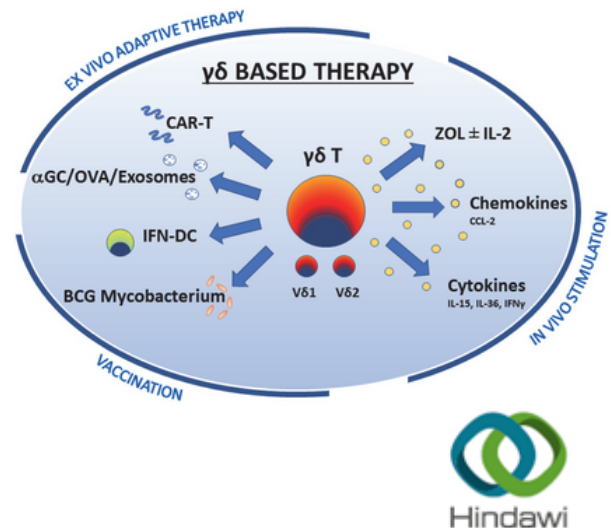
 Γ T Cell-Based Immunotherapy in Melanoma: State of the Art.

F. Toia , A. B. Di Stefano, S. Meraviglia,
E. Lo Presti, R. Pirrello, G. Rinaldi,
F. Fulfaro, F. Dieli, and A. Cordova.

J Oncol. 2019 May 23;2019:9014607. doi:
10.1155/2019/9014607. eCollection
2019.PMID: 31239842

I.F.: 2.60 Q2

Hindawi
Journal of Oncology
Volume 2019, Article ID 9014607, 8 pages
<https://doi.org/10.1155/2019/9014607>

*Review Article* **Γ T Cell-Based Immunotherapy in Melanoma: State of the Art**

F. Toia ,¹ A. B. Di Stefano ,¹ S. Meraviglia,² E. Lo Presti,² R. Pirrello,¹ G. Rinaldi,³
F. Fulfaro,³ F. Dieli,² and A. Cordova ¹

¹Department of Surgical, Oncological and Oral Sciences, Plastic and Reconstructive Surgery Section, University of Palermo, 90127, Italy

²Department of Biopathology, Central Laboratory of Advanced Diagnosis and Biomedical Research (CLADIBIOR), University of Palermo, 90127, Italy

³Department of Surgical, Oncological and Oral Sciences, Oncology Section, University of Palermo, 90127, Italy

Correspondence should be addressed to A. B. Di Stefano; annabarbara.distefano@gmail.com

Received 28 December 2018; Accepted 2 May 2019; Published 23 May 2019

Guest Editor: Aditya B. Pant

Copyright © 2019 F. Toia et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Metastatic melanoma is still associated with a poor prognosis, and there is increasing interest in immunotherapy alone or in combination with other adjuvant therapies. Γ T lymphocytes play a pivot role in the immune response against cancer, but while $\gamma\delta$ -based immunotherapy is already a clinical reality for several solid tumors, data on melanoma are still limited and fragmented. This systematic review presents preclinical and clinical evidence for a role of $\gamma\delta$ T lymphocytes in immunotherapeutic strategies for advanced melanoma and discusses research state of the art and future perspectives. Current strategies focus on in vivo stimulation, and ex vivo adoptive therapy and vaccination; results are promising, but further studies are needed to better investigate the interactions in tumoral microenvironment and to improve clinical efficacy of immunotherapeutic protocols.

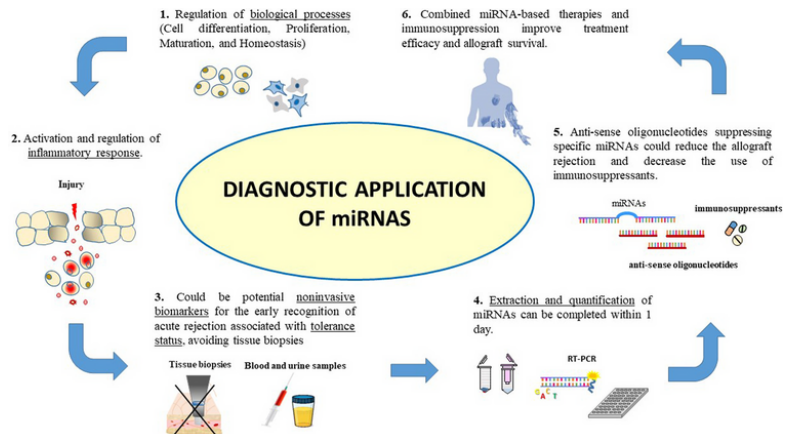
2020

MicroRNAs in solid organ and vascularized composite allotransplantation: Potential biomarkers for diagnosis and therapeutic use.

Anna Barbara Di Stefano, Marco Pappalardo, Francesco Moschella, Adriana Cordova, Francesca Toia.

Transplant Rev (Orlando). 2020
Oct;34(4):100566. doi:
10.1016/j.trre.2020.100566. Epub 2020
Jul 8. PMID: 32682704

I.F.: 3.26 Q1



Transplantation Reviews 34 (2020) 100566



Contents lists available at ScienceDirect

Transplantation Reviews

journal homepage: www.elsevier.com/locate/trre



Review article

MicroRNAs in solid organ and vascularized composite allotransplantation: Potential biomarkers for diagnosis and therapeutic use

Anna Barbara Di Stefano, PhD^{a,1,*}, Marco Pappalardo, MD, PhD^{a,1}, Francesco Moschella, MD^a,
Adriana Cordova, MD^{a,b,c}, Francesca Toia, MD, PhD^{a,b,c}

^a BIOPLAST-Laboratory of Biology and Regenerative Medicine-PLASTIC Surgery, Plastic and Reconstructive Surgery Section, Department of Surgical, Oncological and Oral Sciences, University of Palermo, 90127 Palermo, Italy

^b Plastic and Reconstructive Surgery Section, Department of Surgical, Oncological and Oral Sciences, University of Palermo, 90127 Palermo, Italy

^c Plastic and Reconstructive Unit, Department of Oncology, Azienda Ospedaliera Universitaria Policlinico "Paolo Giaccone", 90127 Palermo, Italy

ARTICLE INFO

Keywords:
Vascularized composite allotransplantation
Micro-RNAs
Biomarker
Immune rejection
Organ transplantation
Tolerance

ABSTRACT

Nowadays, solid organ transplantation (SOT) is an established treatment for patients with end-organ dysfunction, which dramatically improves the quality-of-life. Vascularized composite allotransplants (VCAs) including hand and face have been reported worldwide over the last 20 years. However, VCAs, differently to SOT, are life-enhancing instead of life-saving and are not routinely performed due to the risk of immune rejection and the adverse effects of immunosuppression.

Over the past decade, although considerable improvements in short-term outcomes after allotransplantation have been registered, these results have not been translated into major progress in long-term allograft acceptance and patient survival. Recently active researches in the field of biomarker discovery have been conducted to develop individualized therapies for allograft recipients.

MicroRNAs (miRNAs) are a small noncoding RNAs functioning as critical regulators of gene and protein expression by RNA interference. They have been connected in numerous biological processes and diseases. Due to their immunomodulatory functions, miRNAs have been amended as potential diagnostic and prognostic biomarker for the detection of rejection in allotransplantation. Due to their specific circulating expression profile, they could act as noninvasive predictive tools for rejection that may help clinicians in an early adjustment of the immunosuppression protocol during acute rejections episodes. Indeed, specific anti-sense oligonucleotides suppressing miRNAs expressed in rejection could reduce the rejection rate in allografts and decrease the use of immunosuppressants.

We present a literature review of the immunomodulatory properties and characteristics of miRNAs. We will summarize the current knowledge on miRNAs as potential biomarkers for allograft rejection and possible application in allotransplantation monitoring. Finally, we will discuss the advances in preclinical miRNA-based therapies for immunosuppression.

© 2020 Elsevier Inc. All rights reserved.

2020

Human spheroids from adipose-derived stem cells (S-ASCs) induce calvarial bone production in a xenogeneic rabbit model.

Anna Barbara Di Stefano, Luigi Montesano, Beatrice Belmonte, Alessandro Gulino, Cesare Gagliardo, Ada Maria Florena, Giuseppa Bilello, Francesco Moschella, Adriana Cordova, Angelo A. Leto Barone, Francesca Toia.

Ann Plast Surg. 2020 Dec 15; Publish Ahead of Print. doi: 10.1097/SAP.0000000000002579. Online ahead of print. PMID: 33346554

I.F.: 1.448 Q2

> Ann Plast Surg. 2020 Dec 15; Publish Ahead of Print. doi: 10.1097/SAP.0000000000002579. Online ahead of print.

Human Spheroids from Adipose-Derived Stem Cells Induce Calvarial Bone Production in a Xenogeneic Rabbit Model

Anna Barbara Di Stefano¹, Luigi Montesano, Beatrice Belmonte, Alessandro Gulino, Cesare Gagliardo, Ada Maria Florena, Giuseppa Bilello, Francesco Moschella, Adriana Cordova, Angelo A Leto Barone, Francesca Toia

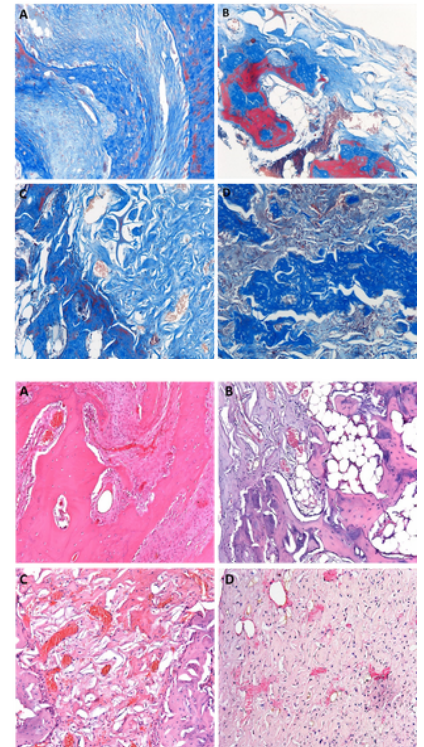
Affiliations + expand

PMID: 33346554 DOI: 10.1097/SAP.0000000000002579

Abstract

Calvarial defects can result from several causes. Tissue engineering hold the potential to restore native form and protective function. We have recently shown that stemness and differentiation ability of spheroids from adipose-derived stem cells (S-ASCs) promotes osteoblasts growth within Integra in a small vertebral lesion. In our study, we aimed to test osteogenic potential of S-ASCs in aiding regeneration of a calvarial defect. Groups containing Integra showed increased bone regeneration at the calvarial defect-Integra interface compared with the control group. In particular, S-ASC-derived osteoblasts group showed a superior calvarial remodeling than undifferentiated S-ASCs group. Clusters of ossification were observed in these both groups with enhanced microvasculature density and fibrosis. In conclusion, seeding of S-ASCs in dermal regeneration templates enhanced bone healing in a rabbit calvarial defect model. These findings could prompt the elective use of S-ASCs with enhanced multilineage differentiation potential for tissue engineering purposes.

Copyright © 2020 Wolters Kluwer Health, Inc. All rights reserved.



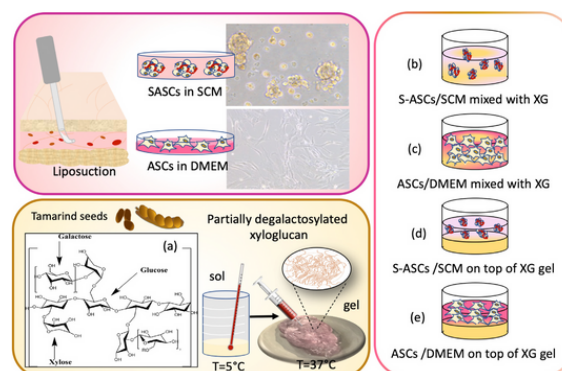
2020

In-situ gelling xyloglucan formulations as 3D artificial niche for adipose stem cell spheroids.

F. Toia, A.B. Di Stefano, E. Muscolino, M.A. Sabatino, D. Giacomazza, F. Moschella, A. Cordova, C. Dispenza. *Int J Biol Macromol.* 2020

Oct 23:S0141-8130(20)34806-6. doi: 10.1016/j.ijbiomac.2020.10.158. Online ahead of print. PMID: 33470202

I.F.: 6.953 Q1



International Journal of Biological Macromolecules 165 (2020) 2886–2899



Contents lists available at ScienceDirect

International Journal of Biological Macromolecules

journal homepage: <http://www.elsevier.com/locate/ijbiomac>



In-situ gelling xyloglucan formulations as 3D artificial niche for adipose stem cell spheroids

F. Toia^{a,b,1}, A.B. Di Stefano^{b,1}, E. Muscolino^c, M.A. Sabatino^c, D. Giacomazza^d, F. Moschella^b, A. Cordova^{a,b}, C. Dispenza^{c,d,*}

^a Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vespro 129, 90127 Palermo, Italy

^b BIOPLAST-Laboratory of BIOlogy and Regenerative Medicine-PLASTic Surgery, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vespro 129, 90127 Palermo, Italy

^c Dipartimento di Ingegneria, Università degli Studi di Palermo, Viale delle Scienze 6, 90128 Palermo, Italy

^d Istituto di BioFisica, Consiglio Nazionale delle Ricerche, Via U. La Malfa 153, 90146 Palermo, Italy

ARTICLE INFO

Article history:
Received 25 June 2020
Received in revised form 16 October 2020
Accepted 20 October 2020
Available online 24 October 2020

Keywords:
Spheroids of adipose stem cells
Artificial niche
In-situ forming gel
Partially degalactosylated xyloglucan

ABSTRACT

Three-dimensional spheroidal cell aggregates of adipose stem cells (SASCs) are a distinct upstream population of stem cells present in adipose tissue, with enhanced regeneration properties *in vivo*. The preservation of the 3D structure of the cells, from extraction to administration, can be a promising strategy to ensure optimal conditions for cell viability and maintenance of stemness potential. With this aim, an artificial niche was created by incorporating the spheroids into an injectable, in-situ gelling solution of partially degalactosylated xyloglucan (dXG) and an ad hoc formulated culture medium for the preservation of stem cell spheroid features. The evolution of the mechanical properties and the morphological structure of this artificial niche was investigated by small amplitude rheological analysis and scanning electron microscopy, respectively. Comparatively, systems produced with the same polymer and the typical culture medium (DMEM) used for adipose stem cell (ASC) growth in adherent cell culture conditions were also characterised. Cell viability of both SASCs and ASCs incorporated inside the hydrogel or seeded on top of the hydrogel were investigated as well as the preservation of SASC stemness conditions when embedded in the hydrogel.

© 2018 Published by Elsevier B.V.

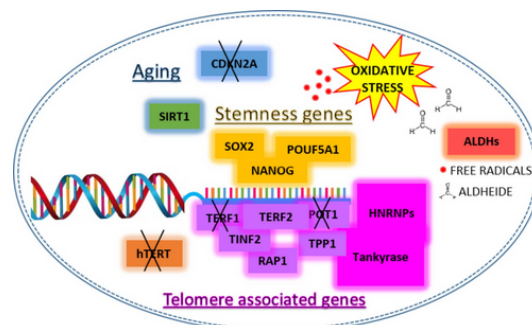
2021

Cell quality evaluation with gene expression analysis of spheroids (3D) and adherent (2D) adipose stem cells.

Anna Barbara Di Stefano, Federica Grisafi, Mileidys Perez-Alea, Marta Castiglia, Marta Di Simone, Serena Meraviglia, Adriana Cordova, Francesco Moschella, Francesca Toia.

Gene. 2021 Feb 5;768:145269. doi: 10.1016/j.gene.2020.145269. Epub 2020 Oct 24. PMID: 33148459

I.F.: 2.984 Q1



Gene 768 (2021) 145269

Contents lists available at ScienceDirect

Gene

journal homepage: www.elsevier.com/locate/gene

Research paper

Cell quality evaluation with gene expression analysis of spheroids (3D) and adherent (2D) adipose stem cells

Anna Barbara Di Stefano^{a,*}, Federica Grisafi^{a,1}, Mileidys Perez-Alea^b, Marta Castiglia^c, Marta Di Simone^{d,e}, Serena Meraviglia^{d,e}, Adriana Cordova^{a,f,g}, Francesco Moschella^a, Francesca Toia^{a,f,g}

^a BIOFLAST-Laboratory of Biology and Regenerative Medicine-FLASTic Surgery, Plastic and Reconstructive Surgery, Department of Surgical, Oncological and Oral Sciences, University of Palermo, 90127 Palermo, Italy

^b Advanced BioDesign, Parc Technologique de Lyon, Woodstock - Bâtiment Cedre 1, Saint Priest, France

^c Medical Oncology, Department of Surgical, Oncological and Oral Sciences, University of Palermo, Palermo, Italy

^d Central Laboratory of Advanced Diagnosis and Biomedical Research (GLADIBIOR), University of Palermo, Palermo, Italy

^e Department of Biomedicine, Neurosciences and Advanced Diagnosis, University of Palermo, Palermo, Italy

^f Plastic and Reconstructive Surgery, Department of Surgical, Oncological and Oral Sciences, University of Palermo, 90127 Palermo, Italy

^g Plastic and Reconstructive Surgery, Department of Oncology, Azienda Ospedaliera Universitaria Policlinico "Paolo Giaccone", 90127 Palermo, Italy

ARTICLE INFO

Keywords:
Adipose stem cells
Spheroid
Aging
Shelterin complex
ALDH family
Telomere length

ABSTRACT

Adipose stem cells (ASCs) represent a reliable source of stem cells with a widely demonstrated potential in regenerative medicine and tissue engineering applications. New recent insights suggest that three-dimensional (3D) models may closely mimic the native tissue properties; spheroids from adipose derived stem cells (SASCs) exhibit enhanced regenerative abilities compared with those of 2D models. Stem cell therapy success is determined by "cell-quality"; for this reason, the involvement of stress signals and cellular aging need to be further investigated. Here, we performed a comparative analysis of genes connected with stemness, aging, telomeric length and oxidative stress, in 3D and 2D primary cultures. The expression levels of stemness-related markers and anti-aging (Sirtuin1) were significantly up-regulated ($P < 0.001$) in SASCs-3D while gene expression of aging-related p16INK4a was increased in ASCs-2D ($P < 0.001$). The 3D and 2D cultures also had a different gene expression profile for genes related to telomere maintenance (Shelterin complex, RNA Binding proteins and DNA repair genes) ($P < 0.01$ and $P < 0.001$) and oxidative stress (aldehyde dehydrogenase class 1 and 3) ($P < 0.05$, $P < 0.01$ and $P < 0.001$) and presented a striking large variation in their cellular redox state. Based on our findings, we propose a "cell quality" model of SASCs, highlighting a precise molecular expression of several genes involved with stemness (SOX2, POU5F1 and NANOG), anti-aging (SIRT1), oxidative stress (ALDH3) and telomeres maintenance.

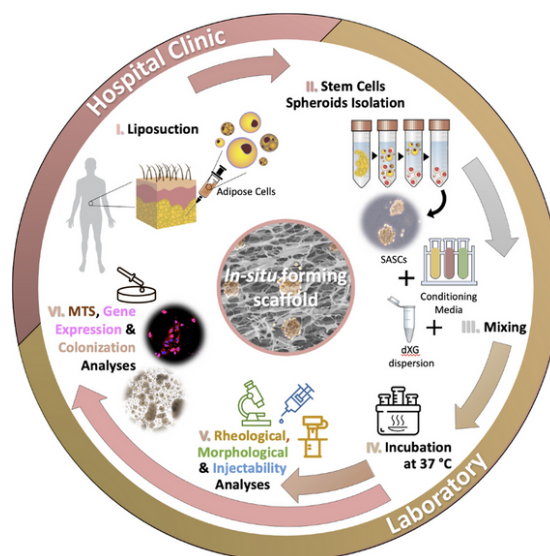
2021

Injectable xyloglucan hydrogels incorporating spheroids of adipose stem cells for bone and cartilage regeneration.

Emanuela Muscolino*,
 Anna Barbara Di Stefano*,
 Marco Trapani, Maria Antonietta Sabatino,
 Daniela Giacomazza,
 Francesco Moschella, Adriana Cordova,
 Francesca Toia,
 Clelia Dispenza.

Materials Science and Engineering: C.
doi.org/10.1016/j.msec.2021.112545.

I.F.: 7.328 Q1



Materials Science & Engineering C 131 (2021) 112545



Contents lists available at ScienceDirect

Materials Science & Engineering C

journal homepage: www.elsevier.com/locate/msec



Injectable xyloglucan hydrogels incorporating spheroids of adipose stem cells for bone and cartilage regeneration

Emanuela Muscolino^a, Anna Barbara Di Stefano^b, Marco Trapani^b, Maria Antonietta Sabatino^a,
 Daniela Giacomazza^c, Francesco Moschella^{b,d}, Adriana Cordova^{b,d}, Francesca Toia^d,
 Clelia Dispenza^{a,c,*}

^a Dipartimento di Ingegneria, Università degli Studi di Palermo, Viale delle Scienze 6, 90128 Palermo, Italy

^b BIOPLAST-Laboratory of Biology and Regenerative Medicine-PLASTic Surgery, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vespere 129, 90127 Palermo, Italy

^c Istituto di Biofisica, Consiglio Nazionale delle Ricerche, Via U. Le Moine 153, 90146, Palermo, Italy

^d Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vespere 129, 90127 Palermo, Italy

ARTICLE INFO

Keywords:

Spheroids of adipose stem cells
 Xyloglucan
 Injectable hydrogels
 Osteogenic differentiation
 Chondroblastic differentiation

ABSTRACT

Cartilage or bone regeneration approaches based on the direct injection of mesenchymal stem cells (MSCs) at the lesion site encounter several challenges, related to uncontrolled cell spreading and differentiation, reduced cell viability and poor engrafting. This work presents a simple and versatile strategy based on the synergic combination of in-situ forming hydrogels and spheroids of adipose stem cells (SASCs) with great potential for minimally invasive regenerative interventions aimed to treat bone and cartilage defects. Aqueous dispersions of partially degalactosylated xyloglucan (dXG) are mixed with SASCs derived from liposuction and either a chondroinductive or an osteoinductive medium. The dispersions rapidly set into hydrogels when temperature is brought to 37 °C. The physico-chemical and mechanical properties of the hydrogels are controlled by polymer concentration. The hydrogels, during 21 day incubation at 37 °C, undergo significant structural rearrangements that support cell proliferation and spreading. In formulations containing 1%w dXG cell viability increases up to 300% for SASCs-derived osteoblasts and up to 1000% for SASCs-derived chondrocytes if compared with control 2D cultures. The successful differentiation into the target cells is supported by the expression of lineage-specific genes. Cell-cell and cell-matrix interactions are also investigated. All formulations resulted injectable, and the incorporated cells are fully viable after injection.

2022

Correlation between tissue-harvesting method and donor-site with the yield of spheroids from adipose-derived stem cells.

Di Stefano AB, Cammarata E, Trapani M, Pirrello R, Montesano L, Meraviglia S, Moschella F, Cordova A, Toia F.

J Plast Reconstr Aesthet Surg. 2022 Sep;75(9):3628-3651.
doi: 10.1016/j.bjps.2022.06.091.
Epub 2022 Jul 22.

I.F.: 4.73 Q1



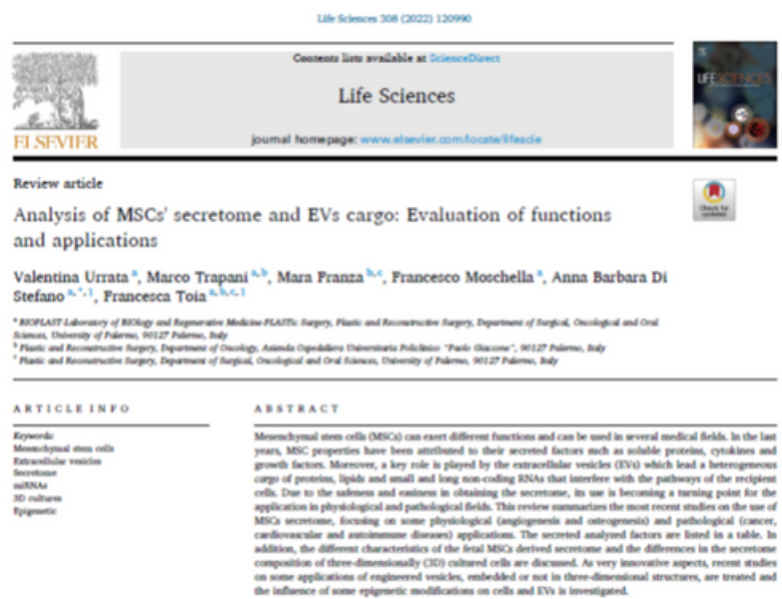
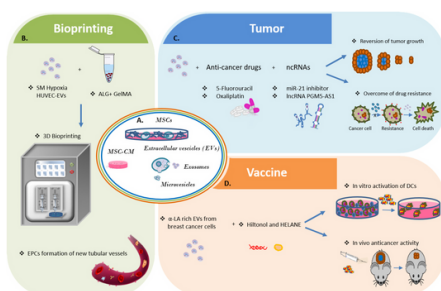
2022

Analysis of MSCs' secretome and EVs cargo: Evaluation of functions and applications.

Urrata V, Trapani M, Franza M, Moschella F, Di Stefano AB, Toia F.

Life Sci. 2022 Nov 1;308:120990.
doi: 10.1016/j.lfs.2022.120990.
Epub 2022 Sep 22.

I.F.: 6.78 Q1

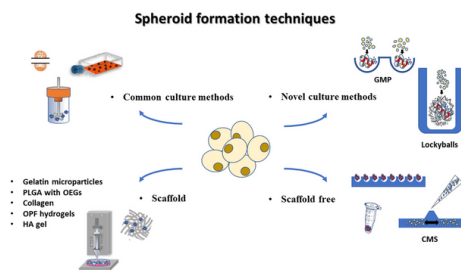


2022

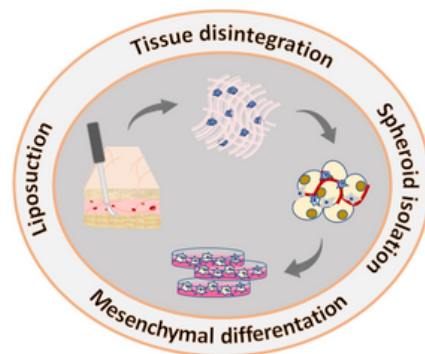
Systematic review on spheroids from adipose-derived stem cells: Spontaneous or artefact state?

Di Stefano AB, Urrata V, Trapani M, Moschella F, Cordova A, Toia F. *J Cell Physiol*. 2022 Oct 9. doi: 10.1002/jcp.30892.

I.F.: 6.513 Q1



Proposed model of Spheroid isolation



Received: 11 February 2022 | Accepted: 22 September 2022
DOI: 10.1002/jcp.30892

REVIEW ARTICLE

Cellular Physiology WILEY

Systematic review on spheroids from adipose-derived stem cells: Spontaneous or artefact state?

Anna Barbara Di Stefano¹ | Valentina Urrata¹ | Marco Trapani¹ |
Francesco Moschella¹ | Adriana Cordova^{1,2,3} | Francesca Toia^{1,2,3}

¹BIORAST-Laboratory of Biology and Regenerative Medicine-PLASTIC Surgery, Plastic and Reconstructive Surgery Unit, Department of Surgical, Oncological and Oral Sciences, University of Palermo, Palermo, Italy

²Department of Surgical, Oncological and Oral Sciences, Unit of Plastic and Reconstructive Surgery, University of Palermo, Palermo, Italy

³Department of D.A.I. Chirurgico, Plastic and Reconstructive Unit, Azienda Ospedaliera Universitaria Polidivisa "Paolo Giaccone", Palermo, Italy

Correspondence
A. B. Di Stefano, University of Palermo, Via
Liborio Guffrè 5, 90127, Palermo, Italy.
Email: annabarbaradiestefano@unipa.it

Funding Information
Progetto Giovani Ricercatori 2016

Abstract

Three-dimensional (3D) cell cultures represent the spontaneous state of stem cells with specific gene and protein molecular expression that are more alike the *in vivo* condition. *In vitro* two-dimensional (2D) cell adhesion cultures are still commonly employed for various cellular studies such as movement, proliferation and differentiation phenomena; this procedure is standardized and amply used in laboratories, however their representing the original tissue has recently been subject to questioning. Cell cultures in 2D require a support/substrate (flasks, multiwells, etc.) and use of fetal bovine serum as an adjuvant that stimulates adhesion that most likely leads to cellular aging. A 3D environment stimulates cells to grow in suspended aggregates that are defined as "spheroids." In particular, adipose stem cells (ASCs) are traditionally observed in adhesion conditions, but a recent and vast literature offers many strategies that obtain 3D cell spheroids. These cells seem to possess a greater ability in maintaining their stemness and differentiate towards all mesenchymal lineages, as demonstrated in *in vitro* and *in vivo* studies compared to adhesion cultures. To date, standardized procedures that form ASC spheroids have not yet been established. This systematic review carries out an in-depth analysis of the 76 articles produced over the past 10 years and discusses the similarities and differences in materials, techniques, and purposes to standardize the methods aimed at obtaining ASC spheroids as already described for 2D cultures.

KEYWORDS

3D cultures, adipose stem cells, biomaterials, hanging drop, spheroids, spinner flask

2022

 κ -Carrageenan and PVA blends as bioinks to 3D print scaffolds for cartilage reconstruction.

Muscolino E*, Di Stefano AB*, Trapani M,
Sabatino MA, Giacomazza D, Alessi S,
Cammarata E, Moschella F, Cordova A, Toia
F, Dispenza C.

Int J Biol Macromol. 2022 Oct 5:S0141-
8130(22)02215-2. doi:
10.1016/j.ijbiomac.2022.09.275.

I.F.: 8.025 Q1



International Journal of Biological Macromolecules 222 (2022) 1861–1875



Contents lists available at ScienceDirect

International Journal of Biological Macromolecules

journal homepage: www.elsevier.com/locate/ijbiomac



κ -Carrageenan and PVA blends as bioinks to 3D print scaffolds for cartilage reconstruction

Emanuela Muscolino^{a,1}, Anna Barbara Di Stefano^{b,1}, Marco Trapani^b,
Maria Antonietta Sabatino^a, Daniela Giacomazza^c, Sabina Alessi^a, Emanuele Cammarata^d,
Francesco Moschella^b, Adriana Cordova^{b,d}, Francesca Toia^{b,d}, Clelia Dispenza^{a,c,*}

^a Dipartimento di Ingegneria, Università degli Studi di Palermo, Viale delle Scienze 6, 90128 Palermo, Italy

^b BIOPLAST Laboratory of BIOlogy and Regenerative Medicine PLASTic Surgery, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vagiro 129, 90127 Palermo, Italy

^c Istituto di Biofisica, Consiglio Nazionale delle Ricerche, Via U. La Malfa 153, 90146 Palermo, Italy

^d Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche, Università degli Studi di Palermo, via del Vagiro 129, 90127 Palermo, Italy

ARTICLE INFO

Keywords:
Spheroids from human adipose stem cells
3D printing
Hydrogel bioinks

ABSTRACT

3D printing of polymeric scaffolds and autologous stem cells is a promising tool for damaged facial cartilage reconstruction surgeries. To this end, suitable bioinks are needed to generate scaffolds with the required morphological and functional features. We formulated hydrogel bioinks using κ -Carrageenan (κ C) and poly(vinyl alcohol) (PVA) in three different weight ratios. The κ C gives the systems the ability to undergo rapid sol-to-gel transitions upon cooling from 60 °C and above to body temperature, while the PVA is used as rheology modifier and porogen. The latter is crosslinked after molding or printing by freeze-thaw cycling for 1 day (FT1) or 5 days (FT5). To select the most suitable formulation for 3D printing, the sol-to-gel transition and the physico-chemical, mechanical and morphological properties of obtained hydrogels were studied. Moreover, the absence of cytotoxic effects of the material on SASCs was assessed in both stemness-preserving or chondro-inductive media. Printing trials were performed to identify optimal process parameters and co-printing and post-printing seeding approaches of SASCs were evaluated. Cells were found to be viable after co-printing and also after the FT1 treatment. Viable adherent cells were also found in the FT5 system, where cells were plated after freezing and thawing treatment.

2022

Re: Correlation between tissue-harvesting method and donor-site with the yield of spheroids from adipose-derived stem cells.

Di Stefano AB, Cammarata E, Trapani M, Pirrello R, Montesano L, Meraviglia S, Moschella F, Cordova A, Toia F.

J Plast Reconstr Aesthet Surg. 2022 Dec 10;77:177-178.

doi: 10.1016/j.bjps.2022.11.055. Epub 2022 Jul 22.

I.F.: 4.73 Q1

Journal of Plastic, Reconstructive & Aesthetic Surgery 77 (2023) 177–178



Correspondence and Communications

Re: Correlation between tissue-harvesting method and donor-site with the yield of spheroids from adipose-derived stem cells

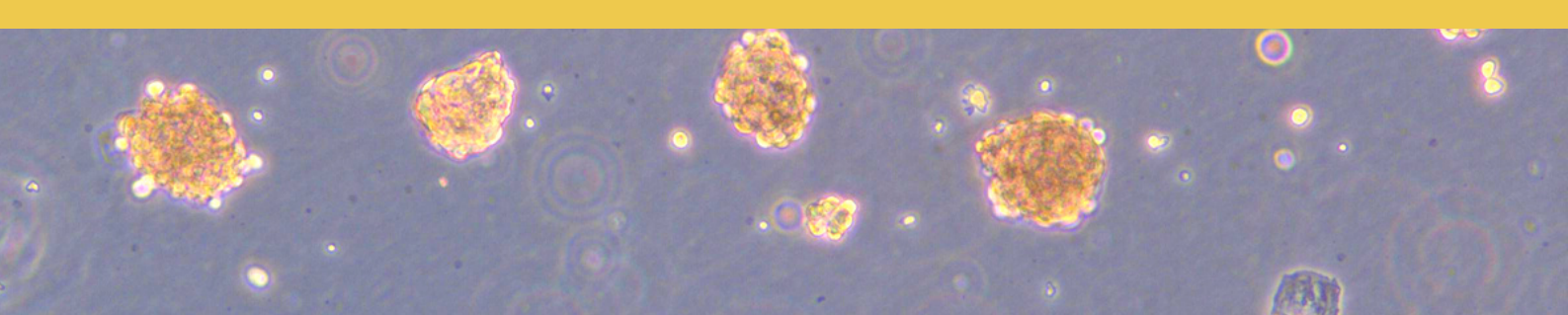


Dear Sir,

We thank Tay et Tay¹ for their reply to our paper, and for giving us the opportunity to focus on the questions and

Table 2 Univariate analysis of the association between age and SASCs density by further stratification in age groups (ANOVA).

Groups	Mean (cells/mL)	F	P-value
15-24	49,899.5	1.5402	0.1768
25-34	39,197.7		
35-44	29,853.0		
45-54	89,823.6		
55-64	23,674.0		
65-74	80,000.0		
75-84	111,111.1		



RELAZIONI A CONVEGNI NAZIONALI E INTERNAZIONALI

1. *Human spheroids from adipose-derived stem cells (s-ascs) induces calvarial bone production in a xenogeneic rabbit model.*

Di Stefano A. B., Montesano L., Maenza A., Belmonte B., Moschella P., Cassata G., Florena A.F., Russo A., Cordova A., Moschella F. and Leto Barone A.A.
Goim 2016

2. *Analysis of gene expression profiles of microRNAs in Spheroids from Adipose-derived Stem Cells (S-ASCs) and their involvement in mesenchymal differentiation and stemness potential.*

A. Barbara Di Stefano, Daniele Fanale, Luigi Montesano, Alessandro Perez, Michele A. Manahan, Justin M. Sacks, Gedge D. Rosson, Antonio Russo, Adriana Cordova, Francesco Moschella and Angelo A. Leto Barone.
IFATS, San Diego (CA) November 17-20 2016.

3. *Involvement of microRNAs in the characterization of mesenchymal differentiation and stemness potential of Spheroids from Adipose-derived Stem Cells (S-ASCs).*

A. Barbara Di Stefano, Daniele Fanale, Luigi Montesano, Alessandro Perez, Antonio Russo, Adriana Cordova, Angelo A. Leto Barone and Francesco Moschella. Regenerative surgery, Roma 2-3 Dicembre 2016.

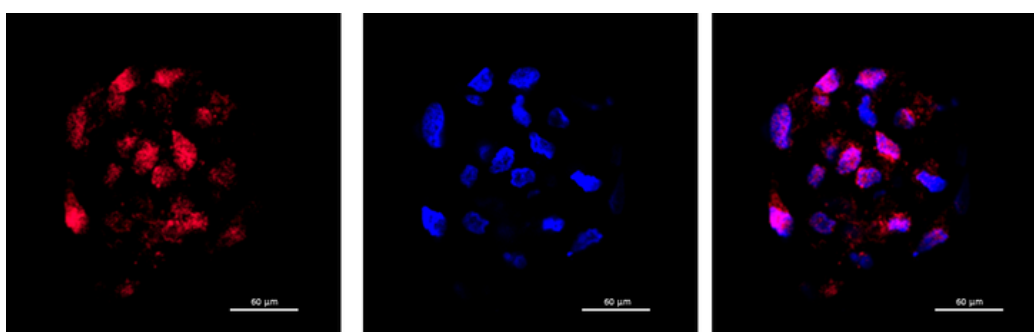
4. *Molecular analysis of Spheroids from Adipose-derived Stem Cells (S-ASCs) during in vitro long-term culture.*

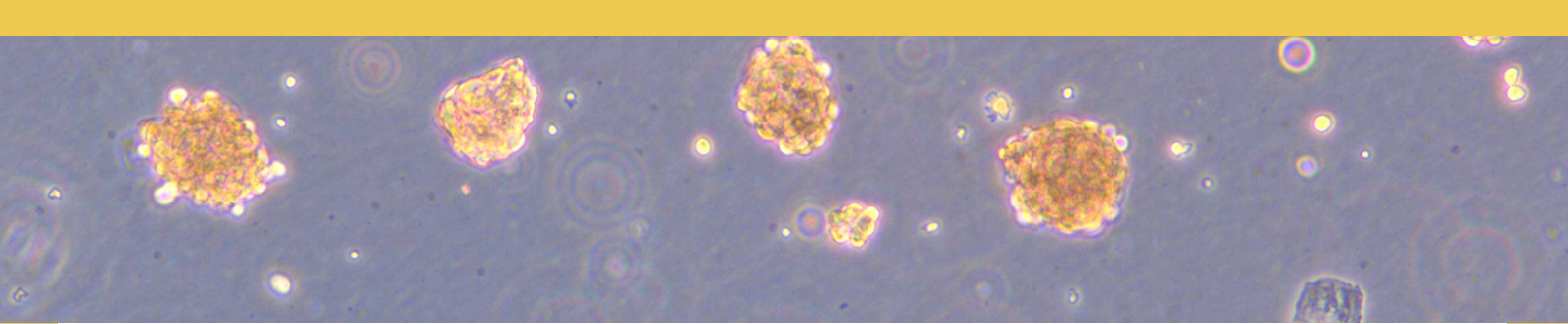
A. Barbara Di Stefano, Federica Grisafi, Alessandro Perez, Marta Castiglia, Antonio Russo, Francesco Moschella, Angelo A. Leto Barone and Adriana Cordova. Regenerative surgery, Roma 15-16 Dicembre 2017.

5. *Capacità rigenerative delle cellule staminali derivate da tessuto adiposo (SASCs) su matrice dermica (INTEGRA®): nostra esperienza su modelli animale.*

Focus su matrici dermiche: Indicazioni, Prospettive e limiti.

Luigi Montesano, Anna Barbara Di Stefano, Moschella Francesco, Adriana Cordova.
Pisa 13/04/2018.





6. *Capacità rigenerative ossee e vascolari degli sferoidi di cellule staminali derivati da tessuto adiposo.*

A. Barbara Di Stefano e F. Moschella
SIMCRI, Taormina 26/10/2018.

7. *Hydrogel scaffolds blends to host Spheroids from human adipose stem cells.*

A.B. Di Stefano, M.A. Sabatino, F. Grisafi, E. Muscolino, M. Perez Alea, F. Toia, C. Dispenza, A. Cordova, F. Moschella.
Regenerative surgery, Roma 21-22 Giugno 2019.

8. *Studio delle proprietà immunomodulatorie degli sferoidi di cellule staminali adipose nei trapianti vascolari compositi.*

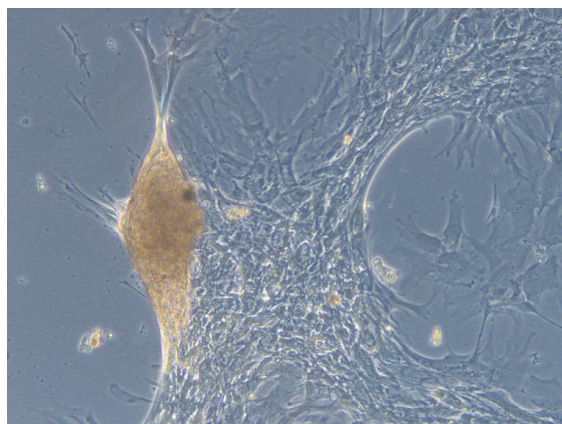
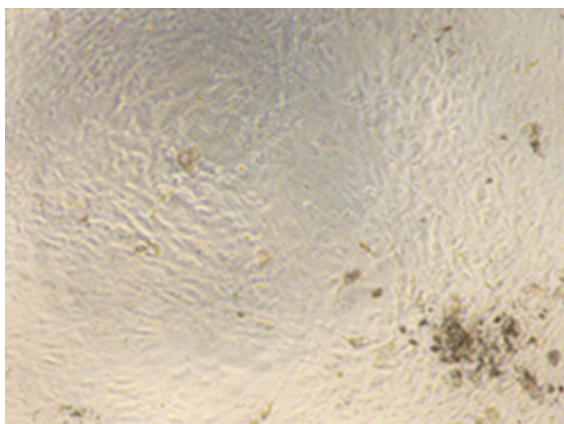
A.B. Di Stefano.
SICPRE, 26-28 Settembre 2019

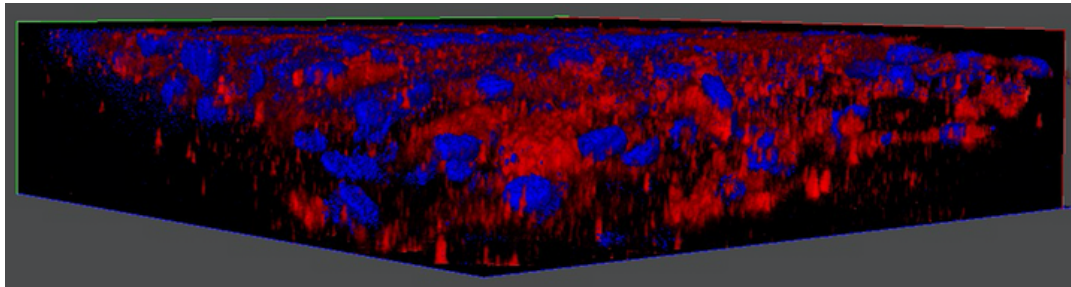
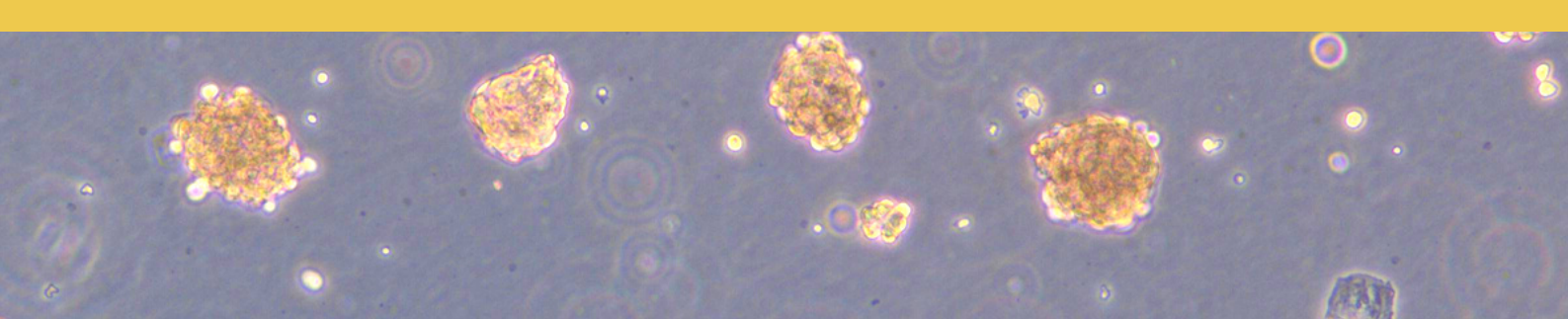
9. *Emerging immunological checkpoints put the brakes on melanoma infiltrating $\gamma\delta$ t cells*

Marta Di Simone, Anna Barbara Di Stefano, Anna Maria Corsale, Luigi Montesano, Elena Lo Presti, Francesca Toia, Adriana Cordova, Francesco Dieli, Serena Meraviglia.
Siica 2021

10. *K-carrageenan/PVA hydrogels as a bio-ink for cartilage reconstruction.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, F. Moschella, A. Cordova, D. Giacomazza C. Dispenza
AERC 2021





11. *Injectable hydrogel formulations to host adipose stem cell spheroids for stemness maintenance and bone and cartilage regeneration.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, F. Moschella, A. Cordova, D. Giacomazza C. Dispenza.

AERC 2021

12. *Miscele di k-carragenina e PVA come bio-inchiostri per stampa 3D di scaffold per la ricostruzione di tessuti cartilaginei.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, M. Trapani, F. Gulino, F. Moschella, A. Cordova, D. Giacomazza, C. Dispenza.

AICling 2021

13. LETTURA: Bioingegneria tissutale: tra presente e futuro.

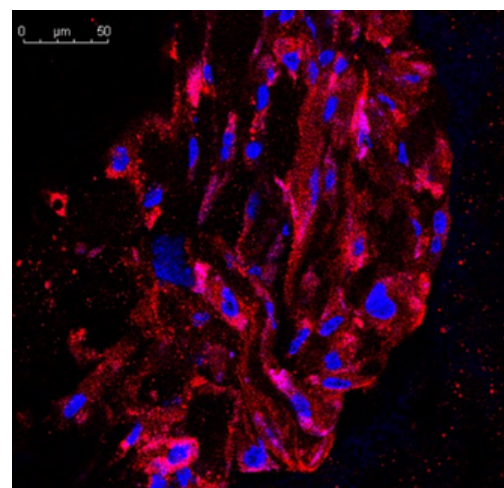
Francesco Moschella.

69° congresso della Società italiana di Chirurgia Plastica. Bologna 24-25 settembre 2021.

14. *Idrogeli con sferoidi di cellule staminali adipose per interventi mini-invasivi di rigenerazione ossea o cartilaginea.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, M. Trapani, F. Moschella, A. Cordova, D. Giacomazza, C. Dispenza.

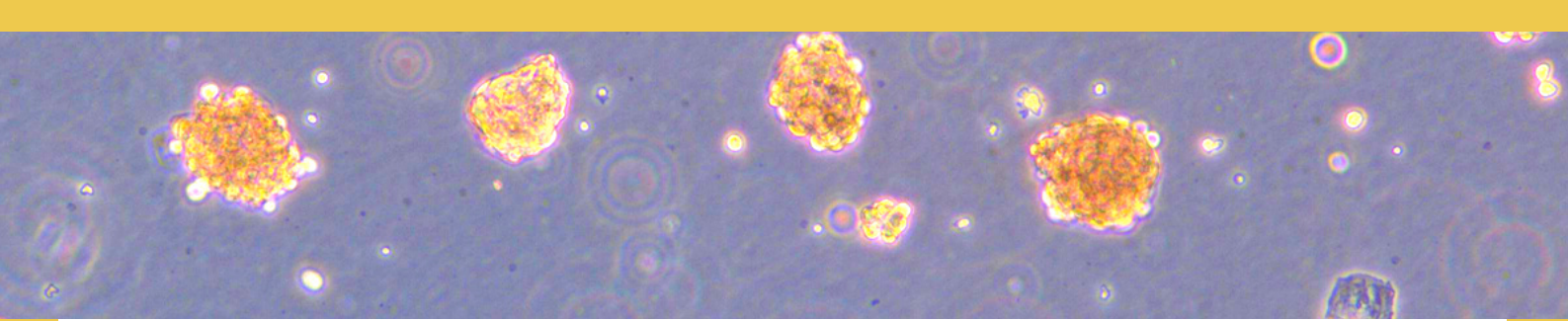
AICling 2021



15. *Adipose stem cell spheroids-laden hydrogels for minimally invasive bone and cartilage regeneration interventions.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, M. Trapani, F. Moschella, A. Cordova, D. Giacomazza, C. Dispenza.

SCI2021



16. *k- Carrageenan and PVA blends as bioinks to 3D print scaffolds for cartilage reconstruction.*

E. Muscolino, A.B. Di Stefano, F. Toia, M.A. Sabatino, M. Trapani, F. Moschella, A. Cordova, D. Giacomazza, C. Dispenza.
SCI2021

17. *Smart hydrogels with Spheroids of Adipose stem cells for minimally invasive bone and cartilage regeneration.*

Anna Barbara Di Stefano, Marco Trapani, Emanuela Muscolino, Maria Antonietta Sabatino, Daniela Giacomazza, Clelia Dispenza, Francesca Toia, Adriana Cordova, Francesco Moschella.
EURAPS 2022

18. *Idrogeli intelligenti con sferoidi di cellule staminali adipose per la rigenerazione minimamente invasiva di ossa e cartilagini.*

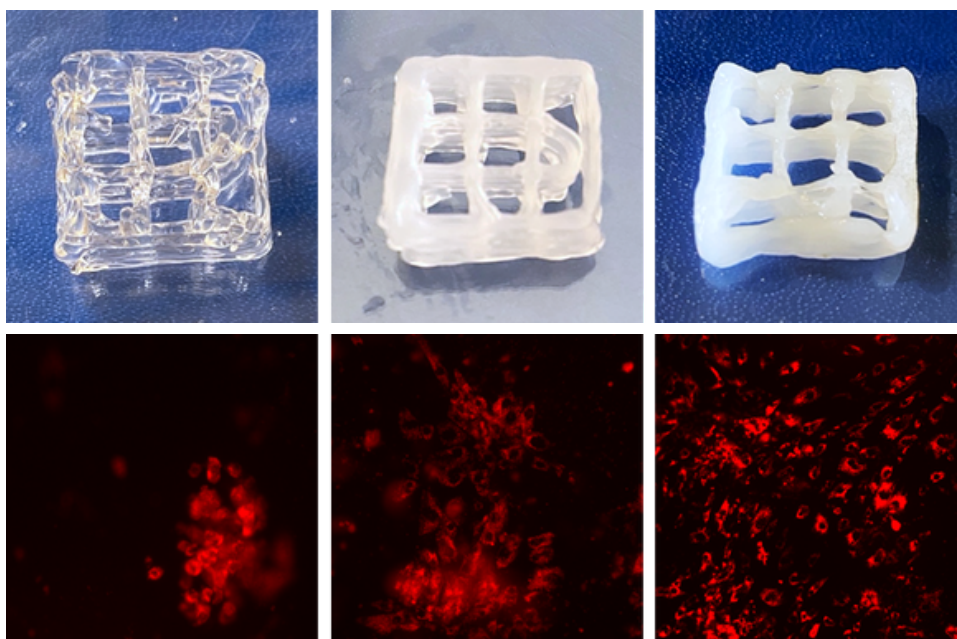
Di Stefano A.B., Trapani M., Muscolino E., Sabatino M.A., Dispenza C., Toia F., Cordova A., Moschella F.
SICPRE 2022

19. *Valutazione delle capacità rigenerative del PRP pediatrico nelle fistole del palato: efficacia, sicurezza e comparazione qualitativa della composizione biochimica.*

Matta D., Pirrello R., Di Stefano A.B., Tondini G., Di Lorenzo S., Cordova A.
SICPRE 2022

20. *LETTURA MAGISTRALE: Proprietà immunomodulatorie di sferoidi di cellule staminali adipose.*

Francesco Moschella.
Università di Sassari, 11 Settembre 2022.



COLLABORAZIONI

Oncologia medica, Dipartimento di Discipline Chirurgiche, Oncologiche e Stomatologiche Università degli Studi di Palermo;

Central Laboratory of Advanced Diagnosis and Biomedical Research (CLADIBIOR), Dipartimento di Biomedicina Neuroscienze e Diagnostica avanzata, Università degli Studi di Palermo;

Dipartimento di Ingegneria, Università degli Studi di Palermo;

Istituto di BioFisica, Consiglio Nazionale delle Ricerche, Palermo;

Advanced BioDesign, Parc Technologique de Lyon, France.

GRANT

BANDO RICERCA FINALIZZATA 2016 GR-2016-02364931

FINANZIATO € 405.345,00

Titolo del Progetto: Role of Human Spheroids from Adipose-derived Stem Cells (SASCs) in hind-limb vascular composite allotransplantation: can they prevent rejection?

RESEARCH TEAM



Prof. Francesco Moschella

Professore onorario Chirurgia Plastica, Università di Palermo.
Fondatore del Laboratorio Bioplast.

franzmoschella@libero.it
francesco.moschella@unipa.it



Prof.ssa Adriana Cordova

Professoressa Ordinario di Chirurgia Plastica Università di Palermo.
Direttrice della UOC di Chirurgia Plastica presso l'AOU Policlinico Universitario.
Direttrice del DAI Chirurgico presso l'AOU Policlinico Universitario.
Board europeo di Chirurgia Plastica (EBOPRAS).

adriana.cordova@unipa.it



Prof.ssa Francesca Toia

Professoressa Associato di Chirurgia Plastica Università di Palermo.
Direttrice Scuola di Specializzazione Chirurgia Plastica. Ricostruttiva ed Estetica,
Università di Palermo.
Board europeo di Chirurgia della mano (EBHS).

francesca.toia@unipa.it

RESEARCH TEAM



Dott.ssa Anna Barbara Di Stefano

Laureata in Scienze Biologiche, Università di Palermo, Anno 2002.
Assegnista di ricerca, Università di Milano nel triennio 2002-2005.
Dottorato in Immunofarmacologia, Università di Palermo, Anno 2008-2010.
Assegnista di ricerca, Università di Palermo, Anno 2012-2013, Laboratorio di Fisiopatologia Cellulare e Molecolare, SSD MED-46.
Assegnista di ricerca, Università di Palermo, Anno 2016-2017,
Borsista PONA3_00011, Università di Palermo, Laboratorio di Fisiopatologia Cellulare e Molecolare, SSD MED-46.

annabarbara.distefano@unipa.it



Ph.D Student Valentina Urrata

Laureata in Biotecnologie per l'Industria e per la Ricerca Scientifica, Università di Palermo, Anno 2020.
PhD Student in Oncologia e Chirurgia sperimentali, Università di Palermo.

valentina.urrata@unipa.it



Borsista Marco Trapani

Laureato in Biotecnologie mediche e medicina molecolare, Università di Palermo, Anno 2020.
Specializzando in Biochimica e patologia clinica, Università di Palermo.

marco.trapani01@unipa.it

